Intended Use
For In Vitro Diagnostic Use.
This histological staining reagent is suitable for visualization of glycogen, basement membrane, mucopolysaccharides, and fungi in tissue using DAKO® automated platforms.

Summary and Explanation
Schiff Solution is a mixture of pararosaniline and sodium bisulfite. Oxidation of tissue sections with periodic acid causes the formation of new aldehyde groups. These new aldehyde groups react with the Schiff Solution to form a colorless unstable compound that becomes a stable pink-to-red colored end product by restoring the quinoid chromophor grouping with a thorough rinse in water. DAKO® Hematoxylin is used as a nuclear counterstain.

Reagents Provided
DAKO® Periodic Acid-Schiff (PAS) Stain System is provided packaged for use on the DAKO® Autostainer. Enough reagents are provided for 50 tests. The DAKO® Periodic Acid-Schiff (PAS) Stain System contains the following reagents:

<table>
<thead>
<tr>
<th>Component</th>
<th>Size</th>
</tr>
</thead>
<tbody>
<tr>
<td>Box 1 of 2 Schiff Solution</td>
<td>4 x 11 mL</td>
</tr>
<tr>
<td>Box 2 of 2 0.5% Periodic Acid</td>
<td>2 x 11 mL</td>
</tr>
<tr>
<td>DAKO® Hematoxylin</td>
<td>2 x 11 mL</td>
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Materials Required, Not Provided
DAKO® Autostainer (Code No. S3400/S3800) and Special Stains software (v.1.2 or higher)
DAKO® Special Stains Wash Buffer 10X (Code No. SL001)
DAKO® Wash Buffer 10X (Code No. S3006)
DAKO® Silanized (Code No. S3003) or poly-L-lysine coated slides
Positive control tissues to use as process controls
Deionized water
95% alcohol
100% alcohol
Xylene or xylene-substitute
Coverslips
Permanent Mounting Medium (Code No. S3026)
Light microscope

Precautions
1. For In Vitro Diagnostic Use.
2. Use prudent laboratory practices when handling reagents. This includes avoiding unnecessary contact, and using personal protective equipment such as chemical resistant gloves, eye protection, and lab coat.
3. Refer to the Material Safety Data Sheet for additional information.

Storage
Store DAKO® Periodic Acid-Schiff (PAS) Stain in original containers. Periodic acid and DAKO® Hematoxylin must be stored at room temperature (20-25°C). Schiff Solution MUST be stored at 2-8 °C. Do not use after the expiration date stamped on the package. To avoid evaporation, keep the Special Stain reagents capped when not in use. If reagents are stored under any conditions other than those specified in the package insert, the user must validate them.

Reagent Preparation
Ready-to-use reagents. No preparation is required. Allow Schiff Solution to come to room temperature before use.
Specimen Collection

Specimens should be collected according to the guidelines published in the NCCLS document M29-T2, “Protection of laboratory workers from infectious diseases transmitted by blood and tissue.”

Process specimens for formalin fixation and paraffin embedding following procedures according to standard histotechnology practices.³

**Note:** For optimal results, specimens should be cut at 4 to 6 microns. Place cut sections towards the bottom of the glass slide at least ½ cm from the edges of the slide and label.

Positive control tissue should be run simultaneously with patient specimens.

Procedure:

The programming steps and incubation times are pre-programmed on the DAKO® Autostainer as follows:

1. 0.5% Periodic Acid - 10 minutes
2. Rinse
3. Schiff Solution - 10 minutes
4. Blow
5. Schiff Solution - 10 minutes
6. Triple Rinse
7. DAKO® Hematoxylin - 6 minutes
8. Double Rinse + Blow

Use a minimum of 400 µL of reagent per slide. Very large tissue sections may require extra reagent.

Prior to starting a run, deparaffinize slides and rinse in water. Place the reagents in the reagent rack according to the reagent map. Pre-soak slides in a working solution of DAKO® Special Stains Wash Buffer for 5 minutes. Place the slides on the DAKO® Autostainer and begin the run.

After the run is complete, dehydrate the slides through 2 changes of 100% ethanol and clear in 2 changes of xylene or xylene substitute. Coverslip with permanent mounting media.

Remove and properly store all reagents at the completion of the run.

Results

Basement membrane, fungi, glycogen, and mucopolysaccharides: Pink to red

Nuclei: Blue

References